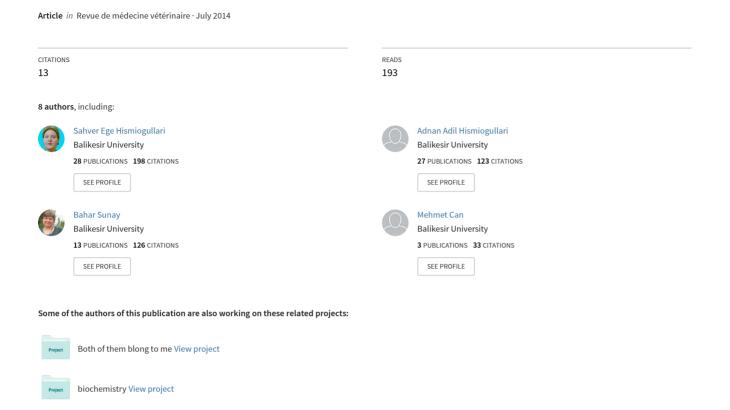
The protective effect of curcumin on carbon tetrachloride induced liver damage



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SUMMARY

Curcumin, a polyphenolic compound of turmeric has been reported to reduce non-alcoholic steatohepatitis (NASH) and oxidative stress in rats. The aim of the present study was to examine the protective effect of curcumin (CUR) on carbon tetrachloride (CCl $_{\rm 4}$) – induced NASH and to evaluate the detailed mechanisms by which CUR exerts its protective action. Thirty male Wistar-Albino rats weighing 250-300 g were randomly divided into three groups: Administrations of oral olive oil (control), CCl $_{\rm 4}$ (0.5 mg/kg in olive oil s.c.) every other day for 3 weeks, and CCl $_{\rm 4}$ (0.5 mg/kg in olive oil s.c. every other day) plus CUR (200 mg/kg/d, orally in olive oil) every day for 3 weeks. The administration of CCl $_{\rm 4}$ resulted in steatohepatitis and increased malondialdehyde (MDA) content of the liver. The administration of CUR decreased lipid deposition during histopathological examination and decreased MDA accumulation. These findings indicate that CUR may have a protective role during liver injury occurring with oxidative damages.

Keywords: Liver fibrosis, steatohepatitis, curcumin, carbon tetrachloride, antioxidant, hepatoprotection

RÉSUMÉ

Effet protecteur de la curcumine sur les dommages hépatiques induits par le tétrachlorure de carbone

La curcumine, est un composé polyphénolique du curcuma précédement signalé comme réduisant la stéatose hépatique non alcoolique (NASH) et le stress oxydatif chez les rats. Le but de la présente étude était d'examiner l'effet protecteur de la curcumine (CUR) sur une hépatite induite par le tétrachlorure de carbone (CCl₄) et d'évaluer les mécanismes par lesquels le CUR exerce son action protectrice. Trente rats albinos Wistar pesant 250-300g ont été répartis au hasard en trois groupes recevant: de l'huile d'olive seule (contrôle), du CCl₄ (0,5 mg/kg dans l'huile d'olive sc) tous les deux jours pendant 3 semaines, et du CCl₄ (0,5 mg/kg dans l'huile d'olive sc tous les deux jours) plus du CUR (200mg/kg/j dans l'huile d'olive oralement) tous les jours pendant 3 semaines. L'administration de CCl₄ a entrainé une stéatose hépatique et augmenté la teneur en malondialdéhyde (MDA). L'administration de la CUR a diminué les dépôts de lipides lors de l'examen histopathologique et diminué l'accumulation de MDA. Ces résultats indiquent que CUR peut avoir un rôle protecteur lors de lésions hépatiques dues à des dommages oxydatifs.

Mots-clés: Fibrose hépatique, stéatose hépatique, curcumine, tetraclorure de carbone, antioxydant, hépatoprotection

Introduction

The molecular mechanism of CCI,-induced hepatotoxicity has been well documented. The trichloromethyl free radical (CCI₃), which is formed in the metabolism of CCI₄ through the cytochrome P450 enzyme system, reacts rapidly with molecular oxygen to produce the trichloromethyl peroxyl radical (CCI₂O₂). This highly toxic radical is responsible for attacks on unsaturated fatty acids of phospholipids present in the cell membrane, leading to lipid peroxidation in the liver cells [45]. Apoptosis also known as programmed cell death, removes the injured cells in the organism without causing any risks to the tissues and organs [37] and during recovery from experimental liver injury, the amounts of activated stellate cells decreases and tissue damage is ameliorated. It has been reported that induction of activated hepatic stellate cells (HSC) apoptosis may cause the elimination of activated HSC for the prevention and treatment of hepatic fibrosis.

Over the years many studies have been conducted investigating the role of antioxidants in experimentally reduced liver damage. We have used the antioxidant curcumin (CUR) which is derived from the plant Curcuma longa (turmeric) and is commonly used in food preparation and is reported to display biological activities such as antioxidant [3], anticarcinogenic [3], anti-inflammatory [13] and immunomodulatory activities [29]. Curcumin has been studied in different models of acute and chronic liver injuries, some of which are alcohol-related or carbon tetrachlorideinduced liver injuries associated with high levels of oxidative stress and inflammation. However, only limited knowledge is available on the possible effects of curcumin on the development and progression of non-alcoholic steatohepatitis (NASH) and is similar to alcoholic steatohepatitis in term of histopatology and characterized by the presence of signs of hepatocellular damage and inflammation, as well as ballooning degeneration and hepatocyte death, formation of Mallory-Denk bodies, and infiltration with inflammatory

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cells [47]. These changes are associated with different degrees of fibrosis or with the presence of cirrhosis. Although some aspects of the pathogenesis of NASH have not been elucidated in detail, others are now well established such as the accumulation of excess lipids particularly in the form of free fatty acids, causes toxic damage to the hepatocytes, which in turn initiates inflammation and tissue repair in the form of fibrosis [9, 25].

The concentration of oxidant and antioxidant components can be measured separately, but the measurement is time-consuming, labor-intensive, costly and requires sophisticated techniques. Due to antioxidant and oxidants molecules have opposite effects, the total antioxidant status [11] (TAS) and the total oxidant status [12] (TOS) can be analyzed by two methods. The present study was conducted to elucidate antioxidative effects of CUR in protecting the hepatotoxic injury caused by CCl, in rats.

Materials and Methods

EXPERIMENTAL GROUPS AND MANAGEMENT

Upon the approval of the experimental protocol by the Animal Ethics Committee of the Balikesir University (Protocol #: 09.11.2010-2010/6-9), animals, obtained from Experimental Animals Breeding and Research Center, Ege University, Turkey, were cared in accordance with National Institutes of Health Guide for the Care and Use of Laboratory Animals. Thirty male Wistar-Albino rats age five months and weighing 250-300 g were housed in separate cages at 25°C and subjected to a 12:12-h light:dark cycle. The rats were randomly divided into three groups: Control group (Group 1), subjected to 1ml olive oil administration by gavage for 3 weeks; CCl₄ group (Group 2), subjected to subcutaneous (sc) CCl₄ (Sigma, St. Louis, MO, USA) injection, 0.5 ml/kg of body weight in olive oil on every other day for 3 weeks, and CCl₄ + CUR group (Group 3), subjected to sc CCl₄ injection, 0.5 ml/kg in olive oil every other day for 3 weeks plus 200 mg/kg of body weight curcumin (Sigma, St. Louis, MO, USA) dissolved in olive oil and given by gavage every day for 3 weeks.

All animals were allowed food $ad \, libitum$ and consumption a standard laboratory diet and had free access to water throughout the experimental period. The rats in all groups were anesthetized on day 21 of the ${\rm CCl_4}$ administration with sodium pentobarbitone (6mg/100 gr of body weight, intraperitoneal). Blood samples, approximately 3 ml from each animal, were collected from abdominal aorta into routine biochemical tubes for biochemical, total antioxidant status and total oxidant status analyses. Then animals were hepatectomized and the liver was separated for further biochemical analysis and histopathological examination.

BIOCHEMICAL ANALYSES

Serum was separated by centrifuging at 825x *g* for 10 min and stored at -80°C for analyses of total protein, total antioxidant status (TAS), total oxidant status (TOS) and as well as alanine aminotransferase (ALT, EC 2.6.1.2), aspartate aminotransferase (AST, EC 2.6.1.1) and alkaline phosphatase (ALP, EC 3.1.3.1) activities using commercially available kits in an auto-analyzer (Cobas Integra 800; Roche Diagnostics GmbH; Mannheim, Germany). Serum and liver protein was measured by Lowry method [22]. The liver tissue measurements were expressed as per gram protein for standardization and accuracy.

PREPARATION OF THE LIVER EXTRACT

After weighing, the liver samples (1 g) were transferred to 9% (w/v) of ice-cold buffered sucrose (0.25 M containing 1 mM HEPES pH 7.4.). The liver was cut into several large pieces and swirled around in the buffer to remove blood as much as possible. The liver was the minced finely with a sharp scissors and transferred to ice-cold homogenizing vessel and was finally homogenized with about six strokes of the pestle at full speed and the homogenate was made up to 10% (w/v) with sucrose buffer solution. A sample of homogenate (3-4 ml) was centrifuged in a fixed angle rotor at 4°C for 10 min at 6000 xg to obtain supernatant for analyses of enzyme, MDA, TAS and TOS. Liver homogenate was stored at -80°C.

DETERMINATION OF TAS AND TOS LEVELS IN SERUM AND LIVER

Serum and liver TAS and TOS levels were measured spectrophometrically (PerkinElmer's Lambda 35 UV/Vis, USA) using commercially available kit (Rel Assay Diagnostic, Turkey). The novel automated method is based on antioxidants in the sample reducing dark blue-green colored 2, 2'-Azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) radical to colorless reduced ABTS form [11]. The change of absorbance at 660 nm is related to total antioxidant level of the sample. The assay is calibrated with a stable antioxidant standard solution which is called as Trolox Equivalent that is a vitamin E analog.

The TOS assay is based on oxidants that are present in the sample and oxidize the ferrous ion chelator complex to ferric ion. The oxidation reaction was prolonged by glycerol molecules abundantly present in the reaction medium. The ferric ion produces a colored complex with chromogen in an acidic medium and the colour intensity, which was measured spectrophotometrically, is related to the total amount of oxidant molecules present in the samples [12]. The assay was calibrated with hydrogen peroxide and the results were expressed in term of micromolar hydrogen peroxide equivalent per liter (µmol $\rm H_2O_2$ Equiv./L) for serum and hydrogen peroxide equivalent per gram protein (µmol $\rm H_2O_2$ Equiv./g protein) for liver extracts.

CALCULATION OF OXIDATIVE STRESS INDEX (OSI)

The ratio of TOS to TAS was accepted as the OSI. In order to calculate this, the resulting unit of TAS was converted to μ mol /L, and the OSI value was calculated regarding to the following formula:

OSI (arbitrary unit) = TOS (μ mol H₂O₂ Equiv./L) / TAS (μ mol Trolox equivalent/L) [48].

DETERMINATION OF SERUM AND LIVER MALON-DIALDEHYTE (MDA) LEVELS

Serum MDA levels were determined based on the spectrophotometric measurement of the product generated upon the reaction of MDA with thiobarbituric acid [37]. The results were expressed as µmol/ml. The MDA content of the homogenates was determined according to the method described by Mihara and Uchiyama [26] and the results were expressed as µmol/g protein.

HISTOPATHOLOGICAL EXAMINATION

The liver specimens were fixed in a 10% neutral buffered formaldehyde solution. After routine processing the tissue specimens were embedded in paraffin wax and sectioned (thickness, $5\,\mu m$). The sections were stained with hematoxylin and eosine (H&E). Histopathological assessment of the H&E- stained section was examined with Olympus BX-51 light microscope.

TUNEL STAINING FOR DETECTION OF APOPTOSIS

Apoptotic cells in the liver sections were detected with the TUNEL staining using a TUNEL assay kit (Millipore, USA) following the manufacturer's instructions.

SCORING AND STATISTICAL ANALYSIS

The sections were scored as described by Noyan et. al. as follows; 0: intact liver; 1: centrilobular necrosis, fatty degeneration; 2: centrilobular and midlobular fatty degeneration, perivenular fibrosis; 3: septal fibrosis, pseudolobule formation; and 4: regenerative nodule formation, cirrhosis [32]. Statistical evaluations were made by using the Mann–Whitney U test and one-way ANOVA after necessary transformation for those data that were not normally distributed (SPSS, version 11.0, Chicago, IL). Group mean differences were attained by the Bonferronipost hoc test and the effect was considered significant when the p value was < 0.05.

Results

BIOCHEMICAL FINDINGS

Serum and hepatic biochemical parameters of rats in control, carbon tetrachloride (CCl_4) and curcumin (CUR) + CCl_4 groups are shown in Table-1.

Administration of CCl₄ increased serum activities of ALT and AST, and serum MDA levels as compared to the control group. Concomitant administration of CUR with CCl₄ suppressed the increase in serum ALT, AST, and MDA levels. Serum TAS decreased while TOS increased in rats administered with CCl₄ as compared to rats in the control group. Serum TAS increased and serum TOS decreased to the control levels when CUR was administered concomitantly with CCl₄. CCl₄ induced elevations in hepatic MDA level and TOS and depression in TAS, which were ameliorated by the CUR treatment (Table-1). CCl₄ increased OSI values both in serum and liver, and CUR when used together with CCl₄, caused a decrease to the control levels (Table-2).

Parameters	Control Group	CCl ₄ Group	CUR + CCl ₄ Group
Serum ALT (IU/L)	120.8±4.4	398.6±12.21 ^{a,*}	128.7±3.7 ^{a,**}
Serum AST (IU/L)	60.7±5.7	190.6±14.2 ^{a,*}	70.2±5.9 ^{a,**}
Serum MDA (μmol/L)	18.3±3.2	53.4±6.4 ^{b,*}	27.2±3.1 ^{b,**}
Serum TAS (mmol Trolox equiv./L)	15.2±0.2	14.1±0.2 ^{b,⋆}	17.3±0.3 ^{b,**}
Serum TOS (μmol H ₂ O ₂ Equiv./L)	32.5±8.2	45.0.5±6.9 ^{b,*}	30.2±5.4 ^{b,**}
Hepatic MDA (μmol/g protein)	8.7 ± 1.4	19.9±2.4 ^{a,*}	11.6±1.03 ^{a,**}
Hepatic TAS (mmol Trolox equiv./g protein)	0.226 ± 0.03	$0.14\pm0.01^{a,*}$	0.212±0.02 ^{a,**}
Hepatic TOS (µmol H ₂ O ₂ Equiv./g protein)	8.7 ± 0.4	12.6±0.8 ^{a,*}	9.1±0.5 ^{a,**}

The data are expressed as mean \pm S.D., (a: P<0.01; b: P<0.001;* compared with control, ** compared with CCl₄), serum aspartate aminotransferase (AST) level, alanine aminotransferase (ALT) level, malondialdehyde (MDA) level, total antioxidant status (TAS) and total oxidant status (TOS) as well as hepatic MDA level, TAS and TOS of rats. Control group received 1 ml olive oil administration by gavage for 3 weeks; CCl₄ group subjected to subcutaneous (sc) CCl₄ injection, 0,5 ml/kg of body weight in olive oil on every other day for 3 weeks, and Cur + CCl₄ group subjected to sc CCl₄ injection, 0,5 ml/kg in olive oil every other day for 3 weeks plus 200 mg/kg of body weight curcumin dissolved in olive oil and given by gavage in every day for 3 weeks.

Table I: Serum and hepatic biochemical parameters of rats in control, carbon tetrachloride (CCl₄) and curcumin (CUR) + CCl₄ groups (n=10 per group).

Groups	Control	CCl	CCl ₄ +CUR
Serum TOS/Serum TAS (arbitrary unit)	19.71	34.65	17.76
Liver TOS/Liver TAS (arbitrary unit)	290	1260	75.83

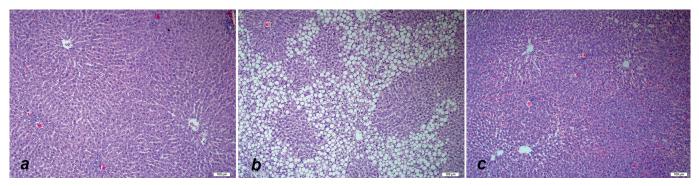


FIGURE 1: Effect of curcumin treatment on rat liver, hematoxylin and eosine. Staining. Representative pictures from the control group (a), CCl_4 group (b) and CCl_4 + Curcumin group (c).

HISTOPATHOLOGICAL FINDINGS

Representative views of liver sections upon CCl₄ induced liver injury and its treatment with CUR is shown in Figure 1. Liver section from the control group showed a normal histological appearance (Figure 1a), whereas livers of rats treated with CCl₄ demonstrated fatty degeneration (Figure 1b). Administration of CUR attenuated fatty degeneration (Figure 1c).

In this study, the severity of fatty degeneration in liver was determined by using a scoring system from 0 to 4. Hepatic damage in $\mathrm{CCl_4} + \mathrm{CUR}$ group was reduced to a median histological score of 0 compared with a median score of 1 in $\mathrm{CCl_4}$ group. In Group 2, $\mathrm{CCl_4}$ caused a significant increase in fatty degeneration scores of animals when compared to control animals, and administration of CUR in $\mathrm{CCl_4} + \mathrm{CUR}$ group caused a significant decrease in this score. There was no significant difference between control group and $\mathrm{CCl_4} + \mathrm{CUR}$ group (Table-3)

DETECTION OF APOPTOTIC CELLS

In this study, TUNEL staining is primarily done for the detection of apoptosis in hepatic satellite cells, owing to the documented apoptotic effect of CUR in these cells [41]. However, there was no difference between study groups neither in apoptosis of hepatic satellite cells, nor hepatocytes (Figure 2).

Discussion

The present study investigated the effect of CUR on oxidative stress markers, TAS, TOS and OSI in conjunction with histopathological and apoptotic analyses of liver in rats treated with ${\rm CCl_4}$. The number of ten rats in each group was designed as the sample size in this study.

The previous experimental studies have shown that treatment with CCl_4 leads to an increase in serum levels of AST, ALT and ALP [2, 24, 31, 40, 43]. The hepatotoxicity of CCl_4 was confirmed in our study by significant increases of

	Morphological scores			
Groups (n=10)	Grade 0	Grade 1	Grade 2	p values
Control	10	-	-	
CCl_4	-	7	3	p<0,0001*
CCl ₄ + CUR	7	3	-	$p < 0.001^{\#} \ p = 0.067^{\phi}$

 ${\rm CCl}_4 = {\rm carbon\ tetrachloride;\ CUR} = {\rm curcumin;\ }^* {\rm\ compared\ with\ control;\ } \# {\rm\ compared\ with\ CCl}_{4;} \oplus {\rm\ compared\ with\ control;\ } \# {\rm\ compared\ with\ control\ with$

TABLE 3: Morphological scores in all groups.

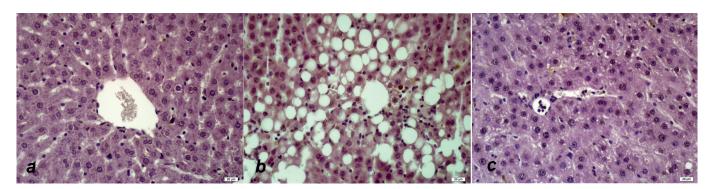


FIGURE 2: Effect of curcumin treatment on the apoptosis of rat liver, TUNEL staining. Representative pictures from the control group (a), CCl₄ group (b) and CCl₄ + Curcumin group (c).

serum levels of AST and ALT. Administration of CUR in rats with hepatic injury, induced by CCl₂ caused a recovery from injury, as evidenced by decrease in the activities of ALT and AST in serum within 3 weeks of treatment. To our knowledge, this study is the first one to determine TAS, TOS and OSI on the protective effects of CUR on CCl,-induced NASH. The major advantage of TAS test is to determine the antioxidant capacity of all antioxidants in a biological sample and not just the antioxidant capacity of a single compound [20]. However, the antioxidant property of curcumin has also been reported by hepatic reduced glutathione (GSH) and the ratio of reduced GSH versus oxidized glutathione (GSSG) parameters. GSH has is a low molecular weight molecule with thiol group and is the most important non-enzyme antioxidant in mammalian cells [13, 45]. Some studies have reported that it inhibitis formation and scavenging of reactive oxygen species (ROS) [1, 10] and reactive nitrogen species [7, 18]. Furthermore, it has been reported that curcumin induces some anti-oxidant enzymes such as such as glutathione transferase [14], haeme-oxygenase-1 [27] and catalase [14]. Taken these observations together, the antioxidant effects curcumin have been studied partly by different researchers but no studies have reported TAS, TOS and ISO.

The findings in the present study consistently support the notion that administration of CUR protects the liver from CCl₄-induced steatohepatitis and injury by attenuating hepatic oxidative stress. These results confirmed that CUR significantly elevated the levels of TAS and OSI in serum and liver extract whilst decreasing levels of TOS and MDA in serum and liver extracts in CCl₄-induced liver injury when compared to CCl₄ group only (Table 1). The results of this study have also demonstrated that CUR significantly reduced pathological changes. Moreover, CUR also significantly decreased the deposition of lipids and normalized the liver changes. HSC presents in the space of Disse and is maintained in a quiescent, non-fibrogenic phenotype in normal liver. It has been reported that curcumin-induced recovery from hepatic injury involves induction of apoptosis of activated the cells [35]. However, there were no significant changes among groups during this duration in apoptosis activities in this rat model (Figure 3). The findings in our study are due to the fact that liver fibrosis did not occur due to the short exposure of the liver to CCl, Priya and Sudhakaran reported that curcumin induced apoptosis only in activated HSCs and not in unactivated HSCs or hepatocytes [35]. In our study, hepatic fibrosis did not occur due to short period of administration CCl₄ to the rats. Therefore, curcumin does not seem to exert apoptosis in CCl₄-induced-NASH for 21 days.

The mechanism of liver steatohepatitis induced by CCl₄ involves peroxidation of the hepatocyte membrane fatty acids which causes destruction of the cell and their intracellular organelles [6]. CCl₄ is activated by liver cytochrome P450 enzymes to form reactive toxic metabolites which can lead to liver injury *in vivo* which is associated with membrane lipid peroxidation and cell necrosis. MDA which

is a stable metabolite of the free radical associated with lipid peroxidation cascade is commonly used as marker of lipid peroxidation. Our results showed that administration of ${\rm CCl_4}$ resulted in a significant increase in MDA production in liver and in serum. The treatment with CUR normalized the increased MDA levels in serum and liver in rats (Figure 1), suggesting that CUR reduced lipid peroxidation in hepatotoxicity, this is in agreements with the results of Basu et al [4]. The study of Park et al [32] also reported similar results that daily administration of 100 mg/kg of curcumin normalized MDA level in acute and subacute rat liver injury induced by ${\rm CCl_4}$.

It has been established that CCl₄ causes necrosis [8, 48], fibrosis [23, 28, 30], mononuclear cell infiltration [37], steatosis and foamy degeneration of hepatocytes, increase in mitotic activity [34] and cirrhosis [31, 43] in liver. Histopathological data also demonstrated alleviation of inflammation and steatosis upon treatment with CUR (Figure 2). Similar results have been reported by Fu et al [13] that curcumin significantly reduces the pathological indexes for hepatocytic death and hepatic injury caused by CCl₄ in the rat model.

Oxidative stress is associated with steatohepatitis [37] and lipid peroxidation and steatohepatitis are significantly attenuated by anti-oxidants such as resveratrol, flavonoid, silymarin or vitamin E. Kowluru et al. [19] reported that the protective effects of curcumin are mediated, in part, by its anti-oxidant defence ability and the scavenging of free radicals; in addition, curcumin is 10 times more active as an anti-oxidant than vitamin E. It has been also shown that CUR decreased the levels of hepatic lipoperoxide formation in both acute and chronic CCl₄ injuries [17, 33]. This alleviation was attributed to CUR's depressive effect on formation of ROS and stimulatory effect on endogenous anti-oxidant activity by its free radical scavenging property [16, 18, 39]. Since ROS and MDA are regarded as inducers of HSC activation [5] the antioxidative properties of CUR may have decreased steatohepatitis in our study (Figure 1). Similarly antioxidant effect of CUR was also shown in rats which developed oxidative stress by thioacetamide administration [38].

In conclusion, the anti-steatohepatitis effect of CUR could be related to its hepatoprotective effects as reflected by reduced MDA and increased TAS through being an antioxidant and exerting anti-inflammatory and free radical scavenger roles, moreover, CUR protects structural and functional integrity of the liver.

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