

# Characterization of essential oils of some *Salvia* species and their antimycobacterial activities

Tülin AŞKUN<sup>1</sup>, K. Hüsnü Can BAŞER<sup>2</sup>, Gülendam TÜMEN<sup>1</sup>, Mine KÜRKÇÜOĞLU<sup>2</sup>

<sup>1</sup>Department of Biology, Faculty of Science and Arts, Balıkesir University, 10145, Balıkesir - TURKEY <sup>2</sup>Department of Pharmacognosy, Faculty of Pharmacy, Anadolu University, 26470 Eskişehir - TURKEY

Received: 02.09.2008

**Abstract:** The compositions of the essential oils of 5 Turkish *Salvia* species, namely *Salvia aucheri* Bentham var. *aucheri* (endemic for Turkey), *Salvia aramiensis* Rech. fil., *Salvia fruticosa* Mill., *Salvia tomentosa* Mill., and *Salvia verticillata* L. subsp. *amasiaca* (Freyn & Bornm.) Bornm., were studied.

Water distilled essential oils from the aerial parts of *Salvia* species from Turkey were analysed by GC and GC/MS. *Salvia* aucheri var. aucheri, *Salvia aramiensis*, and *Salvia fruticosa* oils have the same main constituent: 1,8-cineole (39.2%, 55.6%, and 52.8% respectively).  $\alpha$ -Pinene (25.1%), camphor (14.9%), and borneol (13.2%) were identified as the major components of *Salvia tomentosa*. The main constituents,  $\beta$ -pinene (21.4%) and 1,8-cineole (16.1%), were also the major constituents in the oil of Salvia verticillata subsp. amasiaca.

S. verticillata subsp. amasiaca, S. aucheri subsp. Aucheri, and S. tomentosa showed activity (MIC 196 µg/mL), while S. aramiensis and S. fruticosa did not. This is the first study of the antimycobacterial activity of these 5 plants.

Key words: Antimycobacterial activity, fungi, Salvia, essential oil

# Bazı Salvia türlerinin uçucu yağlarının karakterizasyonu ve antimikobakteriyel aktivitesi

Özet: Türkiyede yetişen 5 Salvia türünün; Salvia aucheri Bentham var. aucheri (Türkiye için endemik), Salvia aramiensis Rech. fil., Salvia fruticosa Mill., Salvia tomentosa Mill. ve Salvia verticillata L. subsp. amasiaca (Freyn & Bornm.) Bornm.'nın uçucu yağ kompozisyonu çalışıldı.

*Salvia* türlerinin topraküstü kısımlarından elde edilen uçucu yağların GC ve GC/MS ile analizleri yapıldı. *Salvia aucheri* var. *aucheri*, *Salvia aramiensis* ve *Salvia fruticosa* yağları ana bileşik olarak 1,8-sineol (sırası ile % 39,2, % 55,6, % 52,8) içermektedir. *Salvia tomentosa* uçucu yağının ana bileşenleri  $\alpha$ -pinene (% 25,1), kafur (% 14,9) ve borneol (% 13,2) olarak bulunmuştur. *Salvia verticillata* subsp. *amasiaca*'nın başlıca bileşenleri  $\beta$ -pinene (% 21,4) ve 1,8-sineol (% 16,1)'dür.

S. *verticillata* subsp. *amasiaca*, S. *aucheri* subsp. *aucheri* ve S. *tomentosa* antimikobakteriyel aktivite gösterirken (MIC 196 μg/mL) S. *aramiensis* ve S. *fruticosa* aktivite göstermedi. Beş *Salvia* türüne ait antimikobakteriyel aktivite çalışması burada ilk defa verilmektedir.

Anahtar sözcükler: Antimikobakteriyel aktivite, fungi, Salvia, uçucu yağ

### Introduction

*Salvia* L. is the largest genus of the family Labiatae, including over 900 species in the world and represented in Turkey by 94 taxa belonging to 89 species with 50% endemism (1,2).

Since ancient times, species of *Salvia* have been used in folk medicine for the treatment of diabetes (3) and skin diseases such as psoriasis and eczema (4). Numerous species of the genus *Salvia* (Labiatae) have been used since ancient times in folk medicine and subjected to extensive pharmacognosic research intended to identify biologically active compounds (5-7).

Salvia species are commonly used in Anatolia for colds, stomach aches, and sore throats. A solution of *Salvia tomentosa* is also used by pouring onto the open cuts and called "Tentürdiyot otu (Iodine tincture herb), "Moşabla" or "Boş yaprak". In addition to *S. fruticosa* tea, called "adaçayı", "elmaçayı" is commonly used to cure colds and stomach aches and other species are used as herbal tea locally (8-10).

*Salvia* species contain various secondary metabolites such as sterols, flavonoids, sesquiterpenoids, sesterpenoids (11), diterpenoids (11-13), triterpenoids (11,14-19), essential oils (13-20), and flavonoids (12).

In a previous study, the essential oils of *S. aucheri* subsp. *aucheri* from a different locality in Turkey were shown to contain  $\alpha$ -pinene (7.6% to 4.3%),  $\beta$ -pinene (6.1% to 4.0%), and 1,8-cineole (39.2% to 20.3%) (24).

Particular interest has been shown in the members of the genus *Salvia* due to a wide range of biological activities such as antifungal activities (25-30), antitumor activities (31-34), antibacterial activities (35-39), antiviral activities (40), cytotoxic activities (41,42), antioxidant activities (36,43), treatment of heart disease (44), and antimycobacterial activity (13).

In addition to these activities, their capability to scavenge free radicals and to inhibit the growth of pathogenic microorganisms (21,45) and antiplatelet aggregation (46), and to inhibit acetyl choline esterase in vitro and in vivo were investigated. The last of these may help explain its traditional use for ailing memory (47,48). *Salvia* species also have some useful compounds to preserve raw and processed food (49) and some of them are used as a drink (50).

To eliminate pathogenic microorganisms, researchers are interested in studying new biologically active compounds isolated from plant species. New studies have shown that some essentials oils could safely be used as antifungal and antibacterial agents to partially or completely inhibit the growth of fungi and bacteria (26,51).

In this study, compositions and antimicrobial activity of the oils of *Salvia aramiensis* Rech. f., *Salvia aucheri* Bentham subsp. *aucheri* (endemic to Turkey), *Salvia fruticosa* Mill., *Salvia tomentosa* Mill., and *Salvia verticillata* L. subsp. *amasiaca* (Freyn & Bornm.) Bornm. were studied. Antimycobacterial activity of the oils is given here for the first time.

Our aim of the study was to determine the major chemicals of the essential oils of *Salvia* species and research their antimycobacterial activity.

#### Materials and methods

## **Plant materials**

Aerial parts of *S. aucheri* subsp. *aucheri*, *S. aramiensis*, *S. fruticosa*, *S. tomentosa*, and *S. verticillata* subsp. *amasiaca* were collected from different parts of Turkey. Locality, altitude, collection time, and herbarium number are given for *Salvia* species in Table 1.

#### Isolation of essential oil

Air-dried aerial parts (90-150 g) were subjected to hydrodistillation for 3 h using a Clevenger-type apparatus to produce the oil. Oil yields are shown in Table 1.

## GC and GC/MS Conditions

The oils were analyzed by capillary GC and GC/MS using an Agilent GC-MSD system.

#### GC/MS

The GC/MS analysis was carried out with an Agilent 5975 GC-MSD system. An Innowax FSC column (60 m  $\times$  0.25 mm, 0.25 m film thickness) was used with helium as carrier gas (0.8 mL/min). GC oven temperature was kept at 60 °C for 10 min and programmed to 220 °C at a rate of 4 °C/min, and kept constant at 220 °C for 10 min and then programmed to 240 °C at a rate of 1 °C/min. Split ratio was adjusted to 40:1. The injector temperature was 250 °C. MS was

Salvia species	Locality	Altitude (m)	Collection date	Oil Yield (%)	Herbarium number
S. aucheri Bentham subsp. aucheri	Mut, Mersin	850 m	13/06/2006	1.8	FS 1543
S. aramiensis Rech. f.	Hatay	350 m	28/06/2006	3.0	FS 1441
S. fruticosa Mill.	Marmara Adası	600 m	01/06/2005	2.3	FS 1423
S. tomentosa Mill.	Kazdagı, Balikesir	850 m	06/07/2006	1.0	FS 1422
S. verticillata L. subsp. amasiaca (Freyn & Bornm.) Bornm	Bitlis, Tatvan, Hizan	1300 m	21/08/2005	0.22	FS 1480

Table 1. Herbarium data of plants and oil yields.

performed at 70 eV. Mass range was from m/z 35 to 450.

#### GC

The GC analysis was carried out using an Agilent 6890N GC system. In order to obtain the same elution order with GC/MS, simultaneous injection was done using the same column and appropriate operational conditions. FID temperature was 300 °C.

The components of essential oils were identified by comparison of their mass spectra with those in the Baser Library of Essential Oil Constituents, Wiley GC/MS Library, Adams Library, and MassFinder Library, and confirmed by comparison of their retention indices. Alkanes were used as reference points in the calculation of relative retention indices (RRIs). Relative percentage amounts of the separated compounds were calculated from FID chromatograms. The results of the analysis are shown in Tables 2-6.

#### Microorganism Used

The oils were tested against the reference strain, *Mycobacterium tuberculosis* H37Ra (ATCC 25177), in duplicate. Inoculums were prepared with 3- to 5-day-old culture of *M. tuberculosis* by diluting 1:5 from MGIT broth, which showed positive.

#### Antimycobacterial Activity

A *Mycobacteria* Growth Indicator Tube (MGIT) containing 4 mL of modified Middlebrook 7H9 Broth Base was used. The assay was done according to the instructions in the MGIT manual fluorometric susceptibility test procedure recommended by the

Table 2. The main compounds of essential oils of *Salvia aucheri* subsp. *aucheri*ç.

RRI	Compounds	%
1032	α-Pinene	7.6
1076	Camphene	7.3
1118	β-Pinene	6.1
1203	Limonene	1.9
1213	1,8-cineole	39.2
1532	Camphor	20.7
1719	Borneol	4.9
2008	Caryophyllene oxide	1.7
2130	Spathulenol	1.1

\* Only the percentages over 1% are indicated in this table.

Table 3. The main compounds of essential oils of Salvia aramiensis.

RRI	Compounds <sup>*</sup>	%
1032	α-Pinene	4.3
1076	Camphene	4.3
1118	β-Pinene	10.2
1174	Myrcene	1.2
1203	Limonene	1.5
1213	1,8-cineole	55.6
1532	Camphor	5.7
1611	Terpinen-4-ol	1.1
1706	α-Terpineol	1.5
1719	Borneol	4.4

\* Only the percentages over 1% are indicated in this table.

RRI

1032

2931

Table 4. The main compounds of essential oils of Salvia fruticosa.

Table 6. The main compounds of essential oils of Salvia verticillata subsp. amasiense.

%

3.3

2.7

Compounds\*

*α*-Pinene

RRI	Compounds*	%
1032	α-Pinene	5.8
1076	Camphene	3.1
1118	β-Pinene	4.5
1174	Myrcene	3.8
1203	Limonene	2.1
1213	1,8-cineole	52.8
1280	<i>p</i> -Cymene	1.4
1437	α-Thujone	1.4
1451	$\beta$ -Thujone	1.1
1532	Camphor	5.8
1612	$\beta$ -Caryophyllene	2.1
1687	α-Humulene	2.6
1706	α-Terpineol	2.1
2008	Caryophyllene oxide	1.1
2104	Viridiflorol	1.1

\* Only the percentages over 1% are indicated in this table.

Table 5. The main compounds of essential oils of Salvia tomentosa.

RRI	Compounds*	%
1032	α-Pinene	25.1
1076	Camphene	4.1
1118	$\beta$ -Pinene	1.6
1174	Myrcene	4.6
1203	Limonene	2.3
1213	1,8-cineole	7.0
1497	α-Copaene	1.0
1532	Camphor	14.9
1590	Bornyl acetate	2.1
1612	β-Caryophyllene	2.2
1687	α-Humulene	2.3
1704	γ-Muurolene	2.6
1719	Borneol	13.2
1773	δ-Cadinene	1.6
2104	Viridiflorol	1.8

\* Only the percentages over 1% are indicated in this table.

β-Pinene 1118 21.4 1132 Sabinene 1.2 Myrcene 1.2 1174 Limonene 1.4 1203 1213 1,8-cineole 16.1 1497 α-Copaene 5.4 1535  $\beta$ -Bourbonene 1.7 1544 α-Gurjunene 4.6 1612  $\beta$ -Caryophyllene 2.3 1661 Alloaromadendrene 5.1 1704 - γ-Muurolene 1.1 1726 Germacrene D 1.2 1755 Bicyclogermacrene 1.6 1773 δ-Cadinene 2.5 2069 Germacrene D-4-ol 1.2 Valeranone 2.5 2145 2187 T-Cadinol 1.2 Copaborneol 1.5 2208 α-Cadinol 2255 2.6

\* Only the percentages over 1% are indicated in this table.

Hexadecanoic acid

manufacturer (Becton Dickinson). OADC enrichment (0.5 mL), a mixture of oleic acid, albumin, dextrose, and catalase, was added to each tube. Oil was added in a volume of 0.1 ml to an MGIT. Then 500  $\mu$ L of bacterial suspension was dispersed in the tubes. The final concentrations of the oil were 196, 98, 49, and 24 µg/mL. An uninoculated MGIT tube was used as a negative control. The control tube contained organisms only and not the oil. Blood Agar was used for checking the growth of other bacteria. The vials were incubated at 37 °C and MIC was determined to be the lowest dilution that gives a negative result by MicroMGIT Fluorescence reader within 2 days when the controls turned positive. Tubes were read daily starting on the second day of incubation using a MicroMGIT Fluorescence reader with a long wave UV light (52).

#### **Results and discussion**

In this study, essential oils of 5 *Salvia* spp., namely *S. aucheri* subsp. *aucheri* (endemic), *S. aramiensis, S. fruticosa, S. tomentosa*, and *S. verticillata* subsp. *amasiaca*, were used (Table 1).

Chemical compositions of these oils were elucidated by GC and GC/MS analysis (Tables 2-6) and the results were evaluated for their in vitro antimycobacterial activity against *M. tuberculosis* (Table 7).

Essential oils of *Salvia species* were screened for antimycobacterial susceptibility testing, and 3 to 5 essential oils, namely *S. aucheri* subsp. *aucheri*, *S. tomentosa*, and *S. verticillata* subsp. *amasiaca*,

Table 7.	Susceptibility	test	results	against	Mycobacterium
	tuberculosis H3	37Ra	(ATCC 2	5177) ob	tained by MGIT
	flourometric m	nanua	l method	•	

Plant no.	Extracts	Concentrations (µg/mL)
1	Salvia aramiensis	n.a.
2	Salvia aucheri subsp. aucheri	196
3	Salvia tomentosa	196
4	Salvia fruticosa	n.a.
5	Salvia verticillata subsp. amasiaca	196
Standard	Streptomycin	0.8
Drugs	Rifampin	1.0
	Ethambuthol	3.5
	Isoniasid	0.1

n.a. not active

#### References

- Davis PH. Flora of Turkey and the Aegean Islands. Edinburgh University Press. Edinburgh; 1982.
- 2. Baser KHC. Aromatic biodiversity among the flowering plant taxa of Turkey. Pure Appl Chem, 74: 527-545, 2002.
- 3. Jimenez J, Risco S, Ruiz T et al. Hypoglycemic activity of *Salvia lavendulifolia*. Planta Med 52: 260-262, 1986.
- Topçu G, Ertaş A, Kolak U, Öztürk M Ulubelen A. Antioxidant activity tests on novel triterpenoids from *Salvia macrochlamys*. ARKIVOC (vii) 195-208, 2007.
- 5. Baytop T. Therapy with medicinal plants in Turkey. University of İstanbul Press. İstanbul; 1984.
- Lu Y, Foo LY. Polyphenolics of Salvia a review. Phytochemistry 59: 114-140, 2002.

exhibited antimycobacterial activity (MIC 196 µg/mL). *S. aramiensis* and *S. fruticosa* were ineffective.

Among the plants that showed antimycobacterial activity, the major components of oils were 1,8-cineole (39.2%), camphor (20.7%),  $\alpha$ -pinene (7.6%), and  $\beta$ -pinene (6.1%) for *S. aucheri* subsp. *aucheri*;  $\alpha$ -pinene (25.1%), camphor (14.9%), borneol (13.2%), and 1,8-cineole (7.0%) for *S. tomentosa*; and  $\beta$ -pinene (21.4%) and 1,8-cineole (16.1%) for *S. verticillata* subsp. *amasiaca*.

The in vitro results obtained in this study provided evidence that some sage oils include chemicals that may have potential as a source of antimycobacterial agents against *M. tuberculosis*.

### Acknowledgements

This study was supported by a grant from the Scientific and Technological Research Council of Turkey (TÜBİTAK), TBAG (Research grant no. 104T336).

Corresponding author: Tülin AŞKUN Department of Biology, Faculty of Science and Arts, Balıkesir University, 10145, Balıkesir - TURKEY E-mail:taskun@balikesir.edu.tr

- Philipson JD. Plants as sources of valuable products. In: Charlwood, BV & Rhodes, MJ (eds) Secondary products from plant tissue culture. Oxford; 1990: pp. 1-22.
- Yeşilada E, Honda G, Sezik E et al. Traditional medicine in Turkey IV, Folk medicine in Mediterranean Subdivision. J. Ethnopharmacol 39: 31-38, 1993.
- Yeşilada E, Honda G, Sezik E et al. Traditional medicine in Turkey V. Folk medicine in the inner Taurus Mountain. J. Ethnopharmacol 46: 133-152, 1995.
- Demirci B, Baser KHC, Tumen G. Composition of the essential oil of *Salvia aramiensis* Rech. Fil. growing in Turkey. J Flav Fragr 17: 23-25, 2002.

- Esquivel B, Sanchez AA, Aranda E. Natural Products of Agricultural Interest from Mexican Labiatae. In: Shahidi F. and Ho CH. eds. Phytochemicals and Phytopharmaceuticals. AOCS Press; 2000: 371-385.
- 12. Ulubelen A, Topcu G. Flavonoids and terpenoids from *Salvia verticillata* and *Salvia pinnata*. J Nat Prod 47:1068-1068, 1984.
- 13. Ulubelen A, Topcu G, Bozok-Johansson C. Norditerpenoids and diterpenoids from *Salvia multicaulis* with antituberculous activity. J Nat Prod 60: 1275-1280, 1997.
- Mehmood S, Riaz N, Nawaz SA. New butyrylcholinesterase inhibitory triterpenes from *Salvia santolinifolia*. Arch Pharm Res 29: 195-198, 2006.
- Pedreros, S. Rodriguez B, de la Torre, MC et al. Dammarane triterpenes of *Salvia hierosolymitana*: structure and absolute stereochemistry of salvilymitone and salvilymitol. Phytochemistry, 29: 919-922, 1990.
- Sokovic M, Tzakou O, Pitarokili D et al. Antifungal activities of selected aromatic plants growing wild in Greece. Die Nahrung 46: 317-320, 2002.
- 17. Ulubelen A, Tan N, Sonmez U et al. Diterpenoids and triterpenoids from *Salvia multicaulis*. Phytochemistry 47: 899-901, 1998.
- Ulubelen A, Oksuz S, Topcu G et al. Antibacterial diterpenes from the roots of *Salvia blepharochlaena*. J Nat Prod 64: 549-551, 2001.
- 19. Ulubelen A, Birman H, Oksuz S Cardioactive diterpenes from the roots of *Salvia eriophora*. Planta Med 68: 818-821, 2002.
- Tabanca N, Demirci B, Baser KHC et al. The chemical composition and antifungal activity of *Salvia macrochlamys* and *Salvia recognita* essential oils. J Agric Food Chem 54: 6593-6597, 2006.
- 21. Tepe, B, Donmez E, Unlu M et al. Antimicrobial and antioxidative activity of the essential oils and methanol extracts of *Salvia cryptantha* (Montbret et Aucher ex Benth.) and *Salvia multicaulis* (Vahl). Food Chem 84: 519-525, 2004.
- Baser K, Kurkcoglu M, Ozek T et al. The Essential Oil of Salvia caespitosa Montbret et Aucher ex Bentham. J Essent Oil Res, 7: 229-230, 1995.
- 23. Kupeli E, Goger F, Kosar M et al. Anti-inflammatory and antinociceptive activities of *Salvia halophila* and *Salvia virgata* from Turkey. Planta Med 73: 836-836, 2007.
- 24. Kurkcuoglu M, Baser KHC, Duman H. Composition of essential oils from two varieties of *Salvia aucheri* Bentham growing in Turkey. J Essent Oil Res, 14: 241-242, 2002.
- 25. Honda G, Koezuka Y, Tabata M. Isolation of an antidermatophytic substance from the root of *Salvia miltirrhiza*. Chem Pharm Bull 36: 408-411, 1988.
- Soliman KM, Badeea RI. Effect of oil extracted from some medicinal plants on different mycotoxigenic fungi. Food Chem Toxicol 40: 1669-1675, 2002.

- Daferera DJ, Ziogas BN, Polission MG. GC-MS analysis of essential oils from some Greek aromatic plants and their fungitoxicity on *Penicillium digitatum*. J Agric Food Chem 48: 2576-2581, 2000.
- Pitarokili D, Tzakou O, Loukis A et al. Volatile metabolites from Salvia fruticosa as antifungal agents in soilborne pathogens. J Agric Food Chem 51: 3294-3301, 2003.
- Viuda M, Ruiz-Navajas Y, Fernández-López J et al. Chemical composition and antifungal activity of the essential oils of *Salvia* (*Salvia officinalis* L.) and rosemary (*Rosemarinus officinalis* L.). Alimentaria 4: 101-105, 2006.
- Gao YG, Song YM, Yang YY et al. Pharmacology of tanshinone. Yao Hsueh Pao 14: 75-82, 1979.
- Chen XG, Li Y, Yan CH et al. Cancer chemopreventive activities of S-3-1, a synthetic derivative of danshione. J Asian Nat Prod Res 3: 63-75, 2001.
- Chaudhuri SK, Badisa RB, Pilarinou E et al. Licamichauxiioic-A and -B acids- two ent-kaurene diterpenoids from *Licania michauxii*. Nat Prod Lett 16: 39-45, 2002.
- 33. Ryu SY, Lee CO, Choi SU. In vitro cytotoxicity of tanshinones from *Salvia* miltiorrhiza. Planta Med 63: 339-342, 1997.
- 34. Topcu G, Tan N, Kokdil G et al. Terpenoids from *Salvia hypargeia*. Phytochemistry 45: 1293-1294, 1997.
- 35. Tzakou O, Pitarokili D, Chinou IB. Composition and antimicrobial activity of the essential oils of *Salvia ringens*. *Planta Med* 67: 81-83, 2001.
- Tepe B, Daferera D, Sokmen A et al. Antimicrobial and antioxidant activities of the essential oil and various extracts of *Salvia tomentosa* Miller (Lamiaceae). Food Chem 90: 333-340, 2005.
- Dobrynin VN, Kolosov MN, Chernov BK et al. Antimicrobial substances of *Salvia officinalis*. Khim Prir Soedin 5: 686-686, 1976.
- Albayrak S, Aksoy M, Hamzaoglu E. Determination of antimicrobial and antioxidant activities of Turkish endemic *Salvia halophila* Hedge, Turk J Biol 32: 265-270, 2008.
- Ogutcu H, Sokmen A, Sokmen M et al. Bioactivities of the various extracts and essential oils of *Salvia limbata* C.A.Mey. and Salvia sclarea L., Turk J Biol 32: 181-192, 2008.
- 40. Watanabe M, Kobayashi Y, Ogihara J et al. HIV-1 reverse transcriptase inhibitory compound in *Salvia officinalis*. Food Sci Tech Res 6: 216-220, 2000.
- Badisa RB, Tzakou O, Couladis M et al. Cytotoxic activities of Salvia plants of the Labiatae family. Pharm Biol 8: 640-645, 2004.
- 42. Guerrero IC, Andres LS, Leon LG et al. Abietane diterpenoids from *Salvia pachyphylla* and *S. clevelandii* with cytotoxic activity against human cancer cell lines. J Nat Prod 12: 1803-5, 2006.
- 43. Weng XC, Wang W. Antioxidant activity of compounds isolated from *Salvia plebeia*. Food Chemistry 71: 489-493, 2000.

- Ginda H, Kusumi T, Ithitsuka MO et al. Salviolone a cytotoxic bisnorditerpene with a benzotropolone chromophore from a Chinese drug Dan-shen (*Salvia miltiorrhiza*). Tetrahedron Lett 29: 4603-4605, 1988.
- Nickavar B, Kamalinejad M, Izadpanah H. In vitro free radical scavenging activity of five *Salvia* species. Pak J Pharm Sci 20: 291-294, 2007.
- 46. Onitsuka M, Fujiu M, Shinma N et al. New platelet aggregation inhibitors from Tan-Shen; radix of *Salvia miltiorrhiza* Bunge. Chem Pharm Bull 31: 1670-1675, 1983.
- 47. Eidi M, Eidi A, Bahar M. Effects of *Salvia officinalis* L. (sage) leaves on memory retention and its interaction with the cholinergic system in rats. *Nutrition* 22: 321-326, 2006.
- Perry NSL, Houghton PJ, Jenner P et al. Salvia lavandulaefolia essential oil inhibits cholinesterase in vivo. Phytomedicine 9: 48-51, 2002.

- 49. Rota C, Carramiñana JJ, Burillo J et al. In vitro antimicrobial activity of essential oils from aromatic plants against selected foodborne pathogens. J Food Prot 67: 1252-1256, 2004.
- 50. Lima CF, Andrade PB, Seabra RM et al. The drinking of a *Salvia officinalis* infusion improves liver antioxidant status in mice and rats. J Ethnopharmacol 2: 383-389, 2005.
- Adam K, Sivropoulou A, Kokkini S et al. Antifungal activities of Origanum vulgare subsp. hirtum, Mentha spicata, Lavandula angustifolia and Salvia fruiticosa essential oil against human pathogenic fungi. J Agric Food Chem, 46: 1739-1745, 1998.
- Becton, Dickinson and Company Newsletter BD Bactec MGIT 960 SIRE kit now FDA-cleared for susceptibility testing of *Mycobacterium tuberculosis*. Microbiology News & Ideas 13: 4-4, 2002.